

The over Heparin 40mL column.

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Spin  $(\text{NH}_4)_2\text{SO}_4$  sol'n - @ 18,000 x g 40 minutes -  
in GSA veta -

Save supernatant -  
Save pellets -

Store one pellet in -20°C - process the other  
pellet 2

pellet 1 slightly greater than half -  $\sim 3/5$

pellet 2 slightly less than half -  $\sim 2/5$

Resuspend pellet in 20 mL of Buffer 1 -

Buffer 1

5 mM Tris pH 7.5

3.1 glycerol

40 mM KCl

5 mM Bme

.1 mM PMSF

dialyze - against Buffer 1 for  $\sim 8$  hrs -

Exchange buffer 4 times -

Heparin column - use prepacked Heparin from A.G. -  
 $\sim 40$  mL column - bump w/ Buffer + KCl -  
wash w/  $\text{H}_2\text{O}$  -

Previously A.G. used 3 mL Heparin a  
5 gram crack

Direct scale up =  $\frac{3}{5} = \frac{4}{5} = 30$  mL Heparin  
 $\sim 50$  g

Equilibrate w/ Buffer 1  $\rightarrow$  (Note: made 20 mM KCl -)

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Read & Understood by me,

Date

Invented by

Date

May Forgo

4/5/95

Recorded by

08/30/95



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Conductivity of Load - 2.8 mS - after ~ 8 hrs of dialysis

Notice a small precipitate matter in dialysis tube -  
Spin down in SS-34 - 18,000 x g - 10 minutes -  
same pellet - small + white -

① Load - 21 mL of sample - 75 ~~mm~~ mL/min - collect FT -

② Wash - 2 V<sub>t</sub> of Buffer 1 - collect <sup>7.5</sup> 8 mL fractions  
1 mL/min

③ Gradient - Buffer 1 to Buffer 2 - 25 mM Tris pH 7.5  
8% glycerol  
5 mM BME  
1 mM PMSF  
2 M KCl

10 V<sub>t</sub> - 400 mL gradient - linear - 1 mL/min -  
collect 7.5 mL fractions -

④ Wash w/ 2 V<sub>t</sub> Buffer 2 1 mL/min - 7.5 mL fractions -

Let column run O/N -

Note Next time gradient should be much shallower - 1 M KCl -

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Witnessed &amp; Understood by me,

Date

Invented by

Date

Mary Long

4/5/95

Recorded by

8/31/95